Changed Fc and C3 Receptor Pattern on Human EBV-negative Lymphoma Cells, Following *in vitro* Conversion by EB-virus*

VIGGO JØNSSON,† GEORGE KLEIN‡ and BJARNE EGELUND CHRISTENSEN†

†Medical-Haematology Department C, Gentofte University Hospital, DK 2900 Hellerup/Copenhagen, Denmark; and ‡Department of Tumor Biology, Karolinska Institutet, S-104 01 Stockholm 60, Sweden

Abstract—EBV-conversion of the originally EBV-negative Ramos and BJAB lymphoma lines has led to an increased frequency of C3 receptor positive and a decreased incidence of Fc receptor positive cells in the population. The concentration of C3 receptors per cell has increased as well. This finding supports the view that EBV-conversion leads to a modification of surface architecture which, in turn, may explain the changes in nutritional and other biological properties.

INTRODUCTION

EPSTEIN—BARR virus (EBV) transforms normal B-lymphocytes into immortal lines that can acquire a certain oncogenic potential. It is important to explore the phenotypic effects of this transformation. In experimental oncogenic (transforming) DNA and RNA virus systems, the study of phenotypic changes at the cell level has given considerable insights into the mechanism of transformation.

One approach to explore the phenotypic consequences of EBV-transformation would be to compare normal, resting B-lymphocytes with EBV-transformed, established However, EBV acts as a polyclonal B-cell activator; it induces immunoglobulin secretion and cellular DNA synthesis [1]. For this between comparisons B-lymphocytes and derived, EBV-transformed lines, while informative, do not specifically focus on the phenotypic changes that are due to the presence of the viral genome as such. Subtle, virally induced changes can be obscured by the massive, pleiotropic activation process. A more sharply focused analysis can be performed by comparing EBV-negative human lymphoma lines of B-cell origin, with their own in vitro EBV-converted sublines.

We have previously described two EBV-negative but EBV-convertible lymphoma

lines, BJAB and Ramos. The former is an exceptional, EBV-negative African Burkitt-like lymphoma [2], while the latter is a typical EBV-negative American Burkitt lymphoma [3]. We have established multiple in vitro EBV-converted sublines from both, by in vitro infection and selection [4–6]. Comparisons between the original BJAB and Ramos lines and their EBV-converted derivatives showed no detectable changes in chromosomal constitution, HLA expression or surface immunoglobulin markers. However, there were profound changes in nutritional and surface characteristics, i.e., increased resistance to saturation conditions, decreased serum dependence, decreased requirements for dialysable serum factors [7-9], decreased capping of various membrane constituents, increased lectine agglutinability [10-12], and increased activation of the alternate complement pathway [13]. The evidence suggests that EBV-conversion is accompanied by important modifications of surface architecture: it is intriguing that they resemble some of the changes associated with classical monolayer transformation by both RNA (Rous, MSV) and DNA (SV40, polyoma, adeno)-viruses.

In order to check the suggestive but largely indirect evidence of membrane rearrangements upon EBV-conversion from a previously unexplored angle, we have examined the expression of two typical lymphocyte markers, Fc and C3 receptors, by a semiquantitative rosetting technique with isotope labelled markers [14]. Pronounced changes were found in the expression of both receptors.

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MATERIALS AND METHODS

Cell lines

The origin and properties of the EBV-negative BJAB and Ramos lymphoma lines were described previously [2, 3]. We have used three EBV-converted, EBNA and EBV-DNA positive sublines of BJAB, BJAB/B95-8, BJAB/HR1K and E95A/BJAB and five EBV-converted sublines of Ramos, namely EHR-A/Ramos, Ramos/B95-8, II-WA-Ramos, AW-Ramos and Ramos/HR1K [5, 15]. All lines were propagated as stationary suspension cultures, on RPMI medium with 10% fetal calf serum.

Measurements of surface Fc and C3 receptors

Cultured cells were washed twice in an 1:1 mixture of Hank's balanced salt solution and isotonic phosphate buffer, pH 7.2 and suspended to a concentration of 4×10^6 ml. No more than 5% of the cells were lost during this procedure. There was no detectable aggregation. Fc and C3 receptors were assayed by the previously described isotope labelled marker cell technique [14]. The procedure can be summarized as follows:

Three test tubes were filled with 1 ml washed cell suspension, adjusted to the same concentration. One millilitre of EA marker cells (viz. sheep erythrocytes sensitized with 1/4 agglutinating titer of rabbit-anti-E-IgGantibodies) was incubated with the target cells in one tube, to estimate the frequency of Fc receptor positive target cells = $L^{\rm Fc}$.

One millilitre of EAC marker cells (viz. sheep erythrocytes sensitized with 1/4 agglutinating titer of rabbit-anti-E-IgM-antibodies plus Complement C3) were incubated with the target cells in the second tube, to estimate the frequency of C3 receptor positive target cell = L^{C3} .

In the third tube, the cells were incubated with 0.5 ml EA and 0.5 ml EAC marker cells, to assess the number of target cells with Fc and/or C3 receptors = $L^{\rm Fc \cup C3}$.

Since Fc and C3 receptors are simultaneously expressed on the surface of some target cells, we have used the previously described formula [14] to compute the number of cells with only Fc receptors = $L^{|\text{Fc}|}$ the number of cells with only C3 receptors = $L^{|\text{C3}|}$ and the number of cells with both Fc and C3 receptors = $L^{\text{Fc} \cap \text{C3}}$.

Using radioactive ⁹⁹Tc labelled EA and ⁵¹Cr labelled EAC marker cells, the mean number of marker cells per rosette was calcu-

lated as a semiquantitative measure of the number of Fc and C3 receptors on $L^{|Fc|}$, $L^{|C3|}$, and $L^{Fc \cap C3}$. Details of this calculation and various control experiments have been described by $J\phi$ nsson and Christensen [14].

RESULTS

As shown in Tables 1 and 2, EBV-converted Ramos and BJAB cells showed an increase in the number of C3-receptors per EAC rosetting cell, both in the $L^{\text{Fc} \cap \text{C3}}$ and on $L^{|\text{C3}|}$ fractions.

There was also a switch, as a rule, from 'non-marginal' to 'marginal' situations, previously defined as a higher number of Fc receptor per $L^{\text{Fc}\cap\text{C3}}$ than per $L^{|\text{Fc}|}$, suggesting a change of the cells from L^{Fc} via $L^{\text{Fc}\cap\text{C3}}$ to L^{C3} [14].

The appearance of marginal situations after conversion implies that the percentage number of L^{Fc} and $L^{|\text{Fc}|}$ decreased in the converted BJAB populations while the mean number of determined EA marker cells per $L^{|Fc|}$ and per $L^{Fc \cap C3}$ decreased in the converted Ramos lines. In two converted sublines, BJAB/B95-8 and EHR-A/Ramos, the changes were so pronounced that the whole $L^{\rm Fc}$ fraction disappeared. In the II-WA-Ramos subline, the same tendency was so strong that we could record the highest number of C3 receptors per $L^{[C3]}$ ever seen corresponding to 46.9 EAC marker cells per $L^{[G3]}$, together with complete obscuring of the $L^{\text{Fc} \cap \text{C3}}$ fraction.

In both EBV negative cell lines the relative frequency of $L^{\rm C3}$ and $L^{\rm [C3]}$ was higher than of $L^{\rm Fc}$ and $L^{\rm [Fc]}$. However, BJAB contained a relatively larger frequency of Fc receptor positive cells and a higher percentage of $L^{\rm Fc \cap C3}$ than Ramos. Ramos had a higher concentration of C3 receptors than BJAB. This may explain why the cells of the converted Ramos sublines had more C3 receptors than the converted BJAB cells.

DISCUSSION

We have shown that EBV conversion has brought about a change in the surface receptor expression of both the BJAB and the Ramos cells. The two lines differed at the point of departure since Ramos showed a higher expression of C3 receptors and BJAB expressed relatively more Fc receptors. In spite of this difference, EBV-conversion changed both cell lines in the same direction,

Table 1. Fc and C3 receptor frequency on BJAB cells and its EBV-converted sublines

			Percen	Percentage of				Number	Number of receptors	ş.	
Target cells	$L^{ m Fc}$	$L^{ m G3}$	$L^{ m Fc}$ $L^{ m C3}$ $L^{ m Fc \cup C3}$ $L^{ m Fc \cap C3}$ $L^{ m Fc }$ $L^{ m C3 }$	$L^{ m Fe\cap G3}$	$L^{ m Fc }$	$T_{[C3]}$	$rac{\mathrm{EA}}{L}^{\mathrm{per}}_{\mathrm{Fc}}$	$\mathop{\mathrm{EA}}_{\mathop{\mathrm{Fc}} \cap \mathrm{C3}}^{\mathop{\mathrm{Per}}}$	$egin{array}{ll} ext{EA} & ext{per} \ L^{ ext{Fe} \cap ext{C3}} \ L^{ ext{Fe} \cap ext{C3}} \end{array}$	$rac{\mathrm{EAC}}{L^{ \mathrm{C3} }}$	Marginal situation
I. EBV -negative $(n=1)$	r(63	0	Ξ	76	r. Cr		y u	, C	0.4	Ž
b)Ab $II. EBV$ -positive	CC	co	/ 0	=	+ 7	76	£:	0.0	7:4	4.3	0
(n=3) BJAB/B95-8	20	86	86	20	0	78	0.0	5.7	10.0	17.2	Yes
BJAB/HR1K	18	72	83	7	11	65	4.0	4.0	10.7	21.6	$^{ m N}_{ m o}$
E95/BJAB	13	95	66	9	7	85	5.2	13.4	11.6	15.9	Yes
Mean	17	87	93	11	9	9/	4.6	7.7	10.8	18.2	
Ratio II mean Imean	0.5	1.4	1.1	1.0	0.3	J.5	9.0	1.4	2.6	4.2	
Suggested effect of EBV conversion	\rightarrow				\rightarrow	←			←	←	No ightarrow Yes

n = number of tests. Marginal situation, see text. Suggested effect: $1.5 \le \text{ratio II}/I \le 0.5$.

Table 2. Fc and G3 receptor frequency on Ramos cells and its converted sublines

			Percentage of	tage of				Number	Number of receptors	S.	:
Target cells	LFc	Γ_{C3}	LFc C3	LFCUC3 LFCUC3	$L^{ \mathrm{Fc} }$ $L^{ \mathrm{C3} }$	$T_{ C3 }$	$EA_{ Fc }$	EAper LFGGG3	$rac{ ext{EAC per}}{L^{ ext{Fc} \cap ext{C3}}} rac{ ext{EAC per}}{L^{ ext{C3} }}$	$rac{ ext{EAC per}}{L^{ ext{C3} }}$	Marginal situation
I. EBV -negative $(n=3)$											
Ramos	8	84	68	3	5	81	16.0	7.8	19.7	99.8	Z
Ramos	Ξ	88	93	9	5	82	13.5	6.11	13.3	91.0	S Z
Ramos	7	81	85	33	4	78	14.7	0.9	18.9	95.8	
Mean	6	84	83	4	.C	80	14.7	8.6	14.7	93.9	2
II. EBV-positive									<u>;</u>	I)	
(n=5)											
EHR-A/Ramos	2	95	95	2	0	93	0.0		37.9	7 64	Voc
Ramos/B95-8	9	96	66	3	က	93	9.0	3.5	27.06	37.8	I GS V
II-WA-Ramos	гC	82	87	0	5	82) 67: i 00:	7:5 0 0	0.0	27.7 46.9	N N
AW-Ramos	12	94	86	8	4	98	4.2	0.00	18.6	31.3	S 2
Ramos/HR1K	÷X5	9/	78	3	2	73	17.3	13.8	25.1	36.8	2 2
Mean	9	83	91	3	33	85	8.0	4.6	22.1	39.1	21
Ratio II mean I mean	0.7	1.1	1.0	8.0	9.0	1:1	0.5	0.5	1.5	1.7	
Suggested effect of EBV conversion							→	\rightarrow	←	←	No ightharpoonup Yes

with a relative increase of C3 receptors and a relative decrease of Fc receptors per cell. This was reflected by the change in the total proportion of receptor positive cells and also by the receptor density per cell.

These observations clearly support the view that EBV-conversion leads to membrane rearrangements. Conceivably, the change in membrane structure may be responsible for the changed nutritional requirements. In Holley's view [16], the changed serum requirements are due to a changed expression of appropriate surface receptors.

The increased expression of C3 receptors may be related to the increased ability of the EBV-converted cell lines to activate the alternate complement pathway [13].

REFERENCES

- 1. A. Rosén, M. Jondal, G. Klein and S. Britton, Polyclonal Ig production after Epstein-Barr virus infection of human lymphocytes *in vitro*. *Nature* (*Lond*.) **267**, 52 (1977).
- 2. J. Menezes, W. Leibold, G. Klein and G. Clements, Establishment and characterization of an Epstein–Barr virus (EBV)-negative lymphoblastoid B-cell line (BJAB) from an exceptional EBV-genome negative African Burkitt's lymphoma. *Biomedicine* 22, 276 (1975).
- 3. G. Klein, B. Giovanella, A. Westman, J. Stehlin and D. Mumford, An EBV-genome negative cell line established from an American Burkitt lymphoma; receptor characteristics. EBV infectability and permanent conversion into EBV-positive sublines by *in vitro* infection. *Intervirology* 5, 319 (1976).
- 4. G. B. CLEMENTS, G. KLEIN and S. POVEY, Production by EBV infection of an EBNA positive subline from an EBNA negative human lymphoma cell line without detectable EBV DNA. *Int.* 7. Cancer 16, 125 (1975).
- 5. G. KLEIN, J. ZEUTHEN, P. TERASAKI, R. HONIG, R. BILLING, M. JONDAL, A. WESTMAN and G. CLEMENTS, Inducibility of the Epstein-Barr virus (EBV) cycle and surface marker properties of EBV-negative lymphoma lines and their *in vitro* converted sublines. *Int. J. Cancer* 18, 639 (1976).
- 6. G. Klein, L. Falk and K. Falk. Antigern-inducing ability of Herpes-virus papio in human and baboon lymphoma lines, compared to Epstein-Barr virus. *Intervirology* **10**, 153 (1978).
- 7. M. STEINITZ and G. KLEIN, Comparison between growth characteristics of an Epstein-Barr virus (EBV)-genome-negative lymphoma line and its EBV-converted subline *in vitro*. *Proc. nat. Acad. Sci.* (Wash.) 72, 3518 (1975).
- 8. M. Steinitz and G. Klein, Epstein-Barr virus (EBV)-induced change in the saturation sensitivity and serum dependence of established EBV-negative lymphoma lines in vitro. Virology 70, 570 (1976).
- 9. M. STEINITZ and G. KLEIN, Further studies on the differences in serum dependence in EBV negative lymphoma lines and their *in vitro* EBV converted, virus-genome carrying sublines. *Europ. J. Cancer* 13, 1629 (1977).
- 10. E. YEFENOF and G. KLEIN, Difference in antibody induced redistribution of membrane IgM in EBV-genome free and EBV positive human lymphoid cells. *Exp. Cell Res.* **99**, 175 (1976).
- 11. E. Yefenof, G. Klein, H. Ben-Bassat and L. Lundin, Differences in the Con A-induced redistribution and agglutination patterns of EBV genome-free and EBV-carrying human lymphoma lines. *Exp. Cell Res.* **108**, 185 (1977).
- 12. E. YEFENOF, G. KLEIN, M. JONDAL and M. B. A. OLDSTONE, Surface markers on human B- and T-lymphocytes. IX. Two-color immunofluorescence studies on the association between EBV receptors and complement receptors on the surface of lymphoid cell lines. *Int. J. Cancer* 17, 693 (1976).
- 13. I. McConnell, G. Klein, T. F. Lint and P. J. Lachmann, Activation of the alternative complement pathway by human B cell lymphoma lines is associated with Epstein-Barr virus transformation of the cells, *Europ. J. Immunol.* **8,** 453 (1978).
- 14. V. Jønsson and B. E. Christensen, Semiquantitative determination of Fc and C3 receptors on human lymphocytes by isotope-labelled marker cells. *Scand. J. Haematol.* **19,** 367 (1977).

- 15. K. O. Fresen and H. Zur Hausen, Establishment of EBNA-expressing cell lines by infection of Epstein-Barr virus (EBV)-genome negative human lymphoma cells with different EBV strains. *Int.* 7. Cancer 17, 161 (1976).
- lymphoma cells with different EBV strains. Int. J. Cancer 17, 161 (1976).

 16. R. W. Holley, R. Armour, J. H. Baldwin, K. D. Brown and Y.-C. Yeh, Density-dependent regulation of growth of BSC-1 cells in cell culture: control of growth by serum factors. Proc. nat. Acad. Sci. (Wash.) 74, 5046 (1977).